

Chemical Inhibitors of the Calcium Entry Channel TRPV6

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ABSTRACT

Purpose Calcium entry channels in the plasma membrane are thought to play a major role in maintaining cellular Ca^{2+} levels, crucial for growth and survival of normal and cancer cells. The calcium-selective channel TRPV6 is expressed in prostate, breast, and other cancer cells. Its expression coincides with cancer progression, suggesting that it drives cancer cell growth. However, no specific inhibitors for TRPV6 have been identified thus far.

Methods To develop specific TRPV6 inhibitors, we synthesized molecules based on the lead compound TH-1177, reported to inhibit calcium entry channels in prostate cancer cells *in vitro* and *in vivo*.

Results We found that one of our compounds (#03) selectively inhibited TRPV6 over five times better than TRPV5, whereas TH-1177 and the other synthesized compounds

preferentially inhibited TRPV5. The IC_{50} value for growth inhibition by blocking endogenous Ca^{2+} entry channels in the LNCaP human prostate cancer cell line was $0.44 \pm 0.07 \mu\text{M}$ compared to TH-1177 ($50 \pm 0.4 \mu\text{M}$).

Conclusions These results suggest that compound #03 is a relatively selective and potent inhibitor for TRPV6 and that it is an interesting lead compound for the treatment of prostate cancer and other cancers of epithelial origin.

KEY WORDS breast cancer · calcium channel · chemical inhibitors · prostate cancer · TRPV6

ABBREVIATIONS

CRAC	Ca^{2+} release-activated Ca^{2+}
FBS	fetal bovine serum
LNCaP	lymph node carcinoma of the prostate
NFAT	nuclear factor of activated T-cells
NMM	N-methylmorpholine
Orai1	calcium release-activated calcium channel protein 1
PyBOP	Benzotriazol-1-yl-oxytripyrrolidinophosphonium
SCID	severe combined immunodeficiency
SOC	store-operated calcium channel
STIM1	stromal interaction molecule 1
THF	Tetrahydrofuran
TRP	transient receptor potential

INTRODUCTION

Prostate cancer is the most common form of cancer in men in the Western world (accounting for one-third of diagnosed cancers) and the second overall leading cause of death from cancer in Europe and the United States. Tumor invasion and metastasis are the primary cause of death (1). At the time of diagnosis, over 70% of men have lymph

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node metastases, which subsequently spread to bone in the pelvic and lumbar vertebral column (2–4). Epidemiologic studies have linked diets rich in calcium to a higher risk of prostate cancer (5–7). However, the biological basis for this link is still unknown. The evidence linking calcium and prostate cancer suggests a critical role for calcium channels in the progression of the disease. Understanding this role may lead to the development of novel therapeutics to improve prostate cancer treatment.

In non-excitabile cell types and some excitable cells, store-operated calcium channels (SOCs) in the plasma membrane serve to replenish intracellular calcium stores in the endoplasmic reticulum following agonist-elicited calcium release (8–10). Besides playing a major role in store-operated calcium entry, these channels are also involved in regulating numerous important physiological processes such as cell growth, proliferation, and apoptosis. The best functionally and biophysically characterized SOC subtypes are the Ca^{2+} release-activated Ca^{2+} (CRAC) channels, originally described in immune cells (11–13) and later in various exocrine cells (e.g., prostate) involved in secretion (14,15). STIM1 and Orai1 have been shown to mediate CRAC currents and store-operated calcium entry in a variety of cells (16–19), but not in prostate cancer cells.

CRAC-like activities have been extensively described for the human prostate carcinoma cell lines PC-3 and LNCaP (14,20). The TRPC1 and TRPV6 members of the transient receptor potential (TRP) ion channel family are the most likely molecular candidates for prostate-specific endogenous SOC or CRAC-like activities (21,22). Numerous studies have linked enhanced endoplasmic reticulum Ca^{2+} accumulation to proliferation in prostate cancer (23–25). Moreover, endogenous SOCs play an important role in the apoptosis of LNCaP prostate cancer cells (26,27).

TRPV6 is strongly expressed in advanced stages of prostate cancer, whereas there is little to no expression evident in healthy tissue and benign prostate hyperplasia (28). TRPV6 transcripts are highly expressed in lymph node metastases originating from the prostate (28), and their expression correlates to the Gleason score and pathological stage (29). There is also evidence that TRPV6 plays a role in breast cancer, being strongly expressed in malignant tissue compared to normal mammary gland tissue (30). We have demonstrated that the breast cancer cell line T47D displays decreased viability after siRNA knockdown of TRPV6 (31).

Several studies have shown that TRPV6 is involved in highly calcium-selective currents in prostate cancer cells and is regulated by intracellular calcium levels (21,32,33). TRPV6-mediated calcium entry has been reported to directly control the proliferation of LNCaP cells and promote apoptosis resistance (34). These studies suggest that TRPV6 would be a promising therapeutic target, although there have been no previous directed efforts to develop inhibitors. The few

known inhibitors include two antifungal compounds econazole and miconazole and the staining dye ruthenium red.

Some evidence that it is possible to target TRPV6 in prostate cancer cells by chemical inhibitors comes from studies conducted by Haverstick *et al.* (35), who developed calcium channel inhibitors structurally similar to dihydropyridines. One of these compounds, TH-1177, effectively blocked store-operated calcium entry in LNCaP prostate cancer cells and inhibited fractional survival of LNCaP cells. When tested in SCID mice carrying prostate cancer cells, TH-1177 treatment prolonged the mean life span of mice by up to 38%. Healthy mice that received 18 times the therapeutic dose showed no signs of toxicity. In these studies, the molecular target of TH-1177 was not identified, although TRPV6 is a likely candidate.

In this study, we make the first directed attempt to create selective inhibitors for TRPV6. Based on the above-mentioned findings, we synthesized compounds that have similar structures to TH-1177 and performed biological assays to screen for TRPV6 inhibition. We identified both TRPV5- and TRPV6-selective inhibitors that were effective at reducing the proliferation of prostate and breast cancer cells. Thus, these inhibitors might be interesting lead compounds in the development of a TRP-targeted cancer treatment.

MATERIALS AND METHODS

Materials

T47D cells and LNCaP cells were obtained from NIH Culture Collection. Plastic six-well cell culture dishes, 96-well cell culture dishes, and T75 cultivation flasks were purchased from BD Biosciences. RPMI-1640 medium from Invitrogen was used to culture the cells. Fetal bovine serum (FBS), trypsin, and penicillin/streptomycin for cell culture, and TRIzol reagent for total RNA purification were also purchased from Invitrogen. TaqMan Universal Master Mix and Primer Express for real-time PCR were from Applied Biosystems. siRNAs and HiPerFect transfection reagent were obtained from Qiagen. Chemicals used for synthesis were from Sigma and Novabiochem.

Cell Culture

The human breast cancer cell line T47D and the human prostate cancer cell line LNCaP were used in this study. Tumor cells were grown in T75 culture flasks in RPMI-1640 medium supplemented with 10% FBS and antibiotics (100 U/ml penicillin and 100 $\mu\text{g}/\text{ml}$ streptomycin) in a humidified atmosphere with 5% CO_2 . Cells used in these studies were between passages 30 to 45. The cells were routinely passaged twice per week.

siRNA Treatment

Two different siRNA molecules were used to knockdown TRPV6 expression in the LNCaP cells. The siRNA1 (target sequence): CTG CAT GTC AGA GCA CTT TAA, siRNA2 (target sequence): AAC CTG CTG CAG CAG AAG AGG, and the siRNA control: AAT CAT CTA AGC TGG CTT TGC were used to transfect into cancer cells. The cells were seeded in 2 ml of culture medium at 400,000 cells per well in six-well plates. After 24 h, siRNA was diluted in phenol red-free medium and delivered with HiPerFect reagent using a final siRNA concentration of 5 nM. The cells were incubated with siRNAs for 72 h before the total RNA was isolated using the TRIzol RNA isolation method, according to the manufacturer's protocol.

Real-Time PCR

cDNA was prepared for each sample by reverse transcription of total RNA using TaqMan reverse transcription reagents according to the manufacturer's manual, as described previously (31). For all experiments, mRNA expression was measured by real-time PCR using an Applied Biosystems 7500 Real-Time PCR System. Reactions consisted of 1x Master Mix, 0.9 μ M forward and reverse primers, and 0.2 μ M dual-labeled fluorescent probes each for TRPV6 and β -actin. The sequences of the forward and reverse primers for TRPV6 were 5'-GGT TCC TGC GGG TGG AA-3' and 5'-CCT GTG CGT AGC GTT GGA T-3', respectively, with the resulting amplicon being 62 bp with a T_m of 60°C. The probe sequence for TRPV6 was 5'-ACA GGC AAG ATC TCA ACC GGC AGC-3'. The forward and reverse primer sequences for β -actin were 5'-CCT GGC ACC CAG CAC AAT-3' and 5'-GCC GAT CCA CAC GGA ATA CT-3', respectively, with the resulting amplicon being 69 bp with a T_m of 60°C. The probe sequence for β -actin was 5'-ATC AAG ATC ATT GCT CCT CCT GAG CGC-3'. Primer Express was used to select primers for TRPV6 and β -actin. All primers were designed to cross exon-exon boundaries of the coding sequence. Primers were optimized and validated for the comparative Ct method, as described in the manufacturer's manual. ABI Prism SDS software version 1 was used for the analysis of the amplification plots. The fold change \pm SD in TRPV6 expression was normalized to β -actin.

Cell Proliferation

Prostate cancer cell proliferation experiments were carried out in 96-well plates. To determine the effect of TRPV6-specific siRNAs on proliferation, 5,000 cells were seeded per well, and after 24 h cells were treated with 5 nM (final concentration) for 72 h. The effect of the synthesized compounds was determined by treating 5,000 cells for 72 h

with log dilutions of the compound (10–0.001 mM). After incubation, the number of living cells was determined using Cell Proliferation Kit II (XTT; Roche) following the manufacturer's instructions. The absorbance was measured by an ELISA reader (Vmax, Molecular Devices) at 450 nm, with a reference wavelength at 650 nm. Data points were measured in triplicates.

TRPV Ca²⁺ Uptake Inhibition

Xenopus oocytes were injected with TRPV5 or TRPV6 cRNA and expressed for 48 h before assaying ⁴⁵Ca²⁺ uptake, as described previously (36). Uptake of ⁴⁵Ca²⁺ was measured in the presence of diluted inhibitor in order to calculate IC₅₀ values.

Ca²⁺ Uptake Activity Measurement

Five-thousand cells per well were seeded in a black 96-well plate in 50 μ l of culture medium per well. After 24 h, siRNA treatment was performed with 5 nM siRNA as described above. Subsequently, after 48 h, the calcium uptake activity was observed in real time using the FLIPR Calcium 3 Assay Kit (Molecular Devices), according to the manufacturer's manual. To the medium, 50 μ l of calcium-free loading buffer containing EGTA (0.1 mM) was added to decrease the influence of FBS in the culture medium. Then 100 μ l of calcium 3 dye was added, and the plate was incubated for 1 h at 37°C. The plate was put into the FlexStation. EGTA (100 mM) was added to the cells to deplete intracellular calcium stores, followed by the addition of calcium solution (100 mM). Fluorescence change (counts) showed the calcium uptake activity after siRNA.

Chemical Synthesis

A general synthesis scheme is shown in Fig. 1. Step 1 yields the amide, step 2 reduces the amide, and step 3 forms the ether bond. Step 1 in synthesis of Compound #01: L-proline methyl ester was coupled with 4-methoxyphenylacetic acid using benzotriazol-1-yl-oxytrypyrrolidinophosphonium (PyBOP) and 2 equivalents of N-methylmorpholine (NMM) to produce methyl 1-[2-(4-methoxyphenyl) acetyl] pyrrolidine-2-carboxylate. In step 2, the amide was reduced to the amino alcohol with LiAlH₄ and AlCl₃ in dry tetrahydrofuran (THF). In step 3, the product was converted to the amine alcohol salt by dissolving the step 2 product in ethyl acetate and 15% HCl/ethyl acetate and evaporating the solvent with a rotary vacuum. This salt was coupled with 4-chlorobenzhydrol (or other alcohol groups as in subsequent syntheses) under Williamson conditions, using catalytic para-toluenesulfonic acid (*p*-TsOH) in refluxing toluene.

The final brownish oil was isolated using column chromatography on silica gel with a 50:50 ethyl acetate:hexane mixture. Structures were confirmed by NMR and mass spectrometry. The three-step synthesis described here provides a general method for synthesis of a large number of derivatives. The following variations were made to the above procedure to synthesize the other compounds. Compound #02: In step 3, benzhydrol was coupled instead of 4-chlorobenzhydrol. Compound #03: In step 3, 4-chlorophenylmethanol was used instead of 4-chlorobenzhydrol. Compound #05: In step 1, 4-dimethylaminophenylacetic acid was used instead of 4-methoxyphenylacetic acid. Compound #06: Same as #05, except the synthesis was ended after step 2. Compound #09: In step 1, 4-chlorophenylacetic acid was used instead of 4-methoxyphenylacetic acid.

Chemical Analysis

Compound #01: purity 90%; ^1H NMR (300 MHz, DMSO- d_6): δ 7.25–7.15 (m, 9H), 6.95–6.82 (d, 2H), 6.65–6.58 (d, 2H), 5.36 (s, 1H), 3.62 (s, 3H), 3.31–3.25 (m, 2H), 2.90–2.80 (m, 2H), 2.69–2.60 (m, 2H), 2.56–2.52 (m, 1H), 2.36–2.30 (m, 2H), 1.77–1.60 (m, 2H), 1.56–1.44 (m, 2H); ESI-MS 436.1 ($\text{M} + \text{H}$) $^+$.

Compound #02: purity 90%; ^1H NMR (300 MHz, DMSO- d_6): δ 7.22–7.18 (m, 10H), 6.97–6.86 (d, 2H), 6.63–6.55 (d, 2H), 5.37 (s, 1H), 3.73 (s, 3H), 3.32–3.24 (m, 2H), 2.94–2.82 (m, 2H), 2.72–2.61 (m, 2H), 2.57–2.54 (m, 1H), 2.37–2.28 (m, 2H), 1.78–1.62 (m, 2H), 1.58–1.44 (m, 2H); ESI-MS 402.1 ($\text{M} + \text{H}$) $^+$.

Compound #03: purity 90%; ^1H NMR (300 MHz, DMSO- d_6): δ 7.41–7.30 (m, 4H), 7.18–7.06 (d, 2H), 6.82–6.79 (d, 2H), 4.47–4.44 (s, 2H), 3.71 (s, 3H), 3.40–3.38 (m, 2H), 2.98–2.85 (m, 2H), 2.64–2.59 (m, 2H), 2.51–2.49 (m, 1H), 2.36–2.28 (m, 2H), 1.79–1.64 (m, 2H), 1.62–1.51 (m, 2H); ESI-MS 360.1 ($\text{M} + \text{H}$) $^+$.

Compound #05: purity 90%; ^1H NMR (300 MHz, DMSO- d_6): δ 7.41–7.32 (m, 9H), 6.94–6.92 (d, 2H), 6.65–6.59 (d, 2H), 5.45 (s, 1H), 3.37–3.33 (m, 2H), 2.92–2.89 (m, 2H), 2.86–2.83 (m, 6H), 2.72–2.65 (m, 2H), 2.57–2.54 (m, 1H), 2.37–2.28 (m, 2H), 1.75–1.63 (m, 2H), 1.58–1.45 (m, 2H); ESI-MS 449.2 ($\text{M} + \text{H}$) $^+$.

Compound #06: purity 95%; ^1H NMR (300 MHz, DMSO- d_6): δ 7.14–7.11 (d, 2H), 6.90–6.81 (d, 2H), 4.30 (s, 1H), 3.39–3.35 (d, 2H), 2.98–2.95 (m, 2H), 2.87–2.83 (m, 6H), 2.68–2.63 (m, 2H), 2.40 (s, 1H), 2.35–2.25 (m, 2H), 1.79–1.62 (m, 2H), 1.59–1.45 (m, 2H); ESI-MS 249.2 ($\text{M} + \text{H}$) $^+$.

Compound #09: purity 90%; ^1H NMR (300 MHz, DMSO- d_6): δ 7.35–7.13 (m, 13H), 5.49 (s, 1H), 3.41–3.34 (m, 2H), 2.98–2.86 (m, 2H), 2.83–2.68 (m, 2H), 2.59–2.54 (m, 1H), 2.34–2.25 (m, 2H), 1.75–1.59 (m, 2H), 1.56–1.44 (m, 2H); ESI-MS 440.1 ($\text{M} + \text{H}$) $^+$.

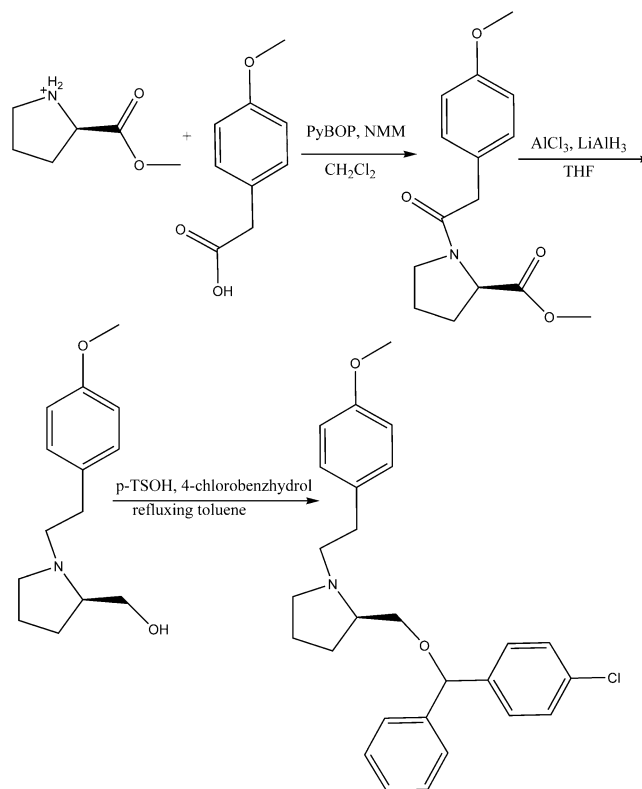


Fig. 1 General scheme for synthesis of chemical compounds.

RESULTS

Chemical Synthesis

To develop inhibitors of TRPV6 we synthesized a group of compounds similar to TH-1177 using a three-step method, as shown in Fig. 1. The structures of the five resulting compounds (#02, #03, #05, #06 and #09) are shown in Fig. 2 and were confirmed by ESI-MS and NMR analysis. Compound #01 is TH-1177. While TH-1177 has been established as an inhibitor of Ca^{2+} entry in prostate cancer cells (35), it has not been previously shown to be a TRPV6 inhibitor. The compounds and controls were tested for their inhibitory potential on TRPV5 and TRPV6, and for their antiproliferative effect in *in vitro* assays using prostate and breast cancer cells.

Inhibition of TRPV5 and TRPV6

To test the potency and selectivity of our compounds for the calcium channels TRPV5 and TRPV6, we performed $^{45}\text{Ca}^{2+}$ uptake assays using TRPV5- or TRPV6-expressing *Xenopus* oocytes. This assay allows for high protein expression and to separately express different isoforms. For each compound and control, IC_{50} values were calculated and are plotted in Fig. 3. The most potent TRPV6 inhibitor was

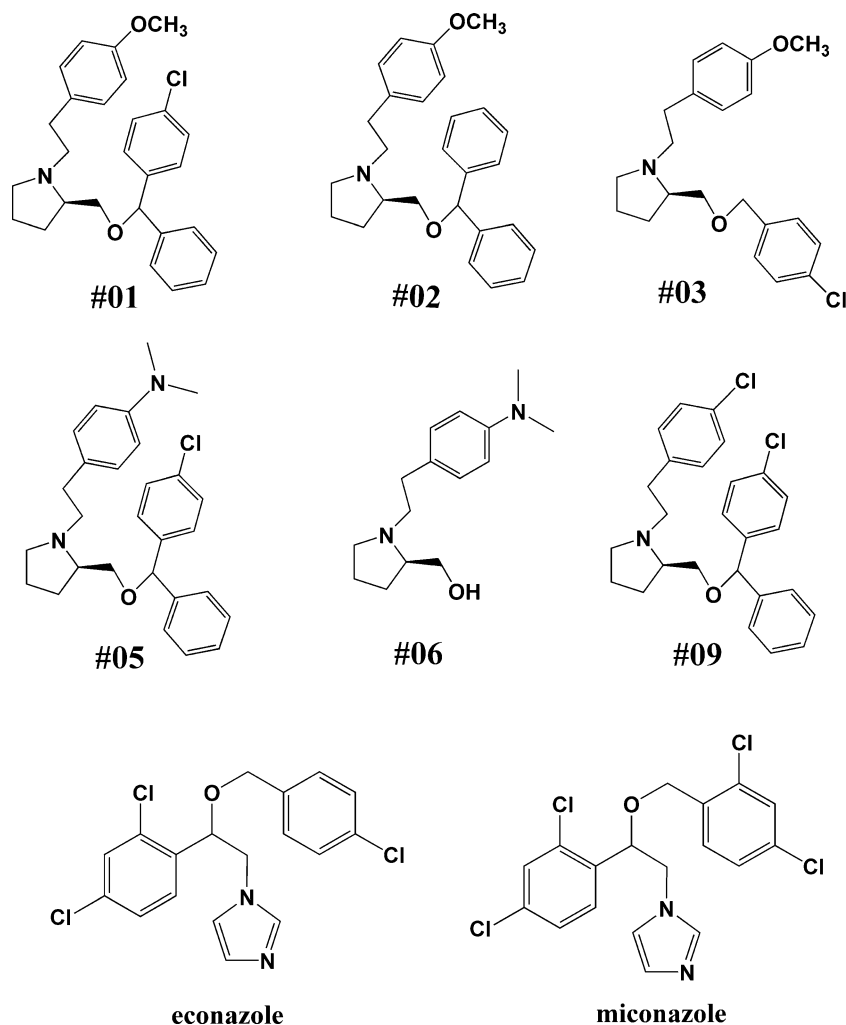


Fig. 2 Structures of synthesized compounds that were tested in *in vitro* assays. Compound #01 is TH-1177. Econazole and miconazole are controls.

compound #03 ($IC_{50}=90 \mu M$), which was over five times less effective at inhibiting TRPV5. Compound #09, which has a chlorinated benzhydryl group, had the strongest inhibitory effect on TRPV5 ($IC_{50}=109 \mu M$), with roughly four times less inhibition of TRPV6. Compound #06, which does not have an ester-linked phenolic side group, did not inhibit either channel and demonstrates that the

presence of an ester-linked side group is critical for activity. Overall, the least effective inhibitors of both channels were compounds #01 and #02. Both control compounds, econazole and miconazole, inhibited TRPV6 more than TRPV5. IC_{50} values and the compound selectivity index (TRPV5/TRPV6) are displayed in Table I and reveal that compound #03 is the most potent and selective for TRPV6.

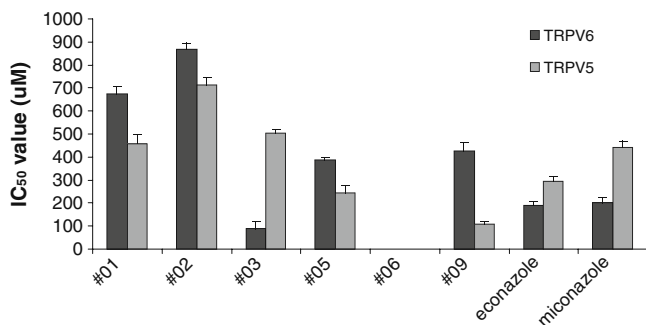


Fig. 3 TRPV-mediated $^{45}Ca^{2+}$ uptake inhibition. IC_{50} values of tested compounds on TRPV5 and TRPV6 expressed in *Xenopus* oocytes.

Growth Inhibition in Cancer Cell Lines

To analyze the effectiveness of our compounds on cancer cell growth, we selected the LNCaP human prostate cancer cell line, since TRPV6 is endogenously expressed and has been reported to play a predominant role in the Ca^{2+} entry pathway in this cell line (21). To confirm this finding, two distinct siRNAs were used to knock down TRPV6 expression in LNCaP cells. In Fig. 4A, the effectiveness of the siRNA treatments can be seen in their ability to reduce TRPV6 mRNA expression. The siRNA1 reduced expression by 55% and siRNA2 by 50%, whereas the control

Table I IC₅₀ Inhibition Values and Selectivity Index (TRPV5/TRPV6) of Compounds Tested in TRPV5- or TRPV6-expressing *Xenopus* Oocytes

Compound	IC ₅₀ TRPV6 (μM)	IC ₅₀ TRPV5(μM)	TRPV5/TRPV6 ^a
#01	675	456	0.68
#02	869	712	0.82
#03	90	503	5.56
#05	386	246	0.64
#06	No effect	No effect	No effect
#09	428	109	0.25
Econazole	190	296	1.56
Miconazole	201	442	2.20

^a Larger (TRPV5/TRPV6) ratios indicate a more TRPV6-selective compound

siRNA treatment did not affect TRPV6 mRNA. This expression knockdown in turn led to a reduction in LNCaP cell proliferation by 57% and 52%, respectively (Fig. 4B). Control siRNA treatment did not affect prostate cancer cell growth.

Next, the synthesized compounds were tested for their ability to inhibit the growth of LNCaP prostate cancer cells as well as a breast cancer cell line endogenously expressing TRPV6. Their effectiveness was similar in

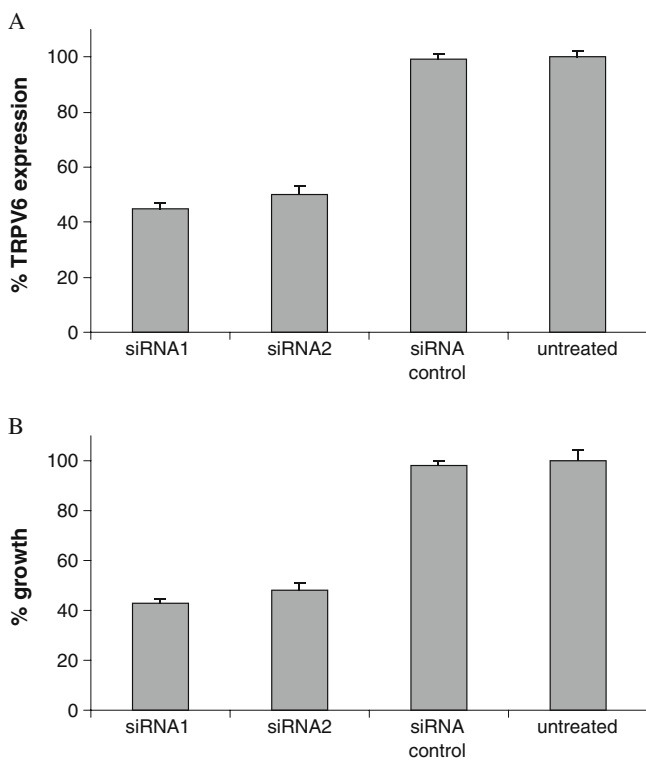


Fig. 4 siRNA knockdown treatments in LNCaP cells. **(A)** Reduction in TRPV6 expression (%) after treatment with TRPV6 siRNAs compared to control siRNA. TRPV6 mRNA level standardized to β-actin. **(B)** Effect of TRPV6 siRNA treatments on cell viability compared to control siRNA, determined by XTT assay.

pattern to the uptake inhibition data in the oocytes. Compound #03 was the best growth inhibitor of T47D cells (IC₅₀=38 μM) and the second best inhibitor of LNCaP cells (IC₅₀=0.44 μM). Likewise, compounds #01 and #02 had the weakest effect in both the prostate and breast cancer cells. Compound #06, which did not show calcium uptake inhibition activity in the oocytes, also showed no effect on reducing cell growth in either cancer cell line (Fig. 5A, B and Table II).

DISCUSSION

Calcium is a central signaling ion that is crucial for controlling growth, proliferation, and survival of normal and malignant cells. Recent studies have revealed that the calcium entry channel TRPV6 is involved in the growth control of prostate cancer cells via Ca²⁺/NFAT-dependent pathways (34). We have also shown that it plays a role in breast cancer, since siRNA knockdown of TRPV6 reduces proliferation in the T47D breast cancer cell line (31).

In this study, we created the first set of chemical compounds directed at TRPV6 inhibition and demonstrated their inhibitory potential in *in vitro* studies. Compound #01, TH-1177, was previously shown to inhibit Ca²⁺ influx into LNCaP human prostate cancer cells (35). However, the molecular target for TH-1177 in LNCaP cells was not

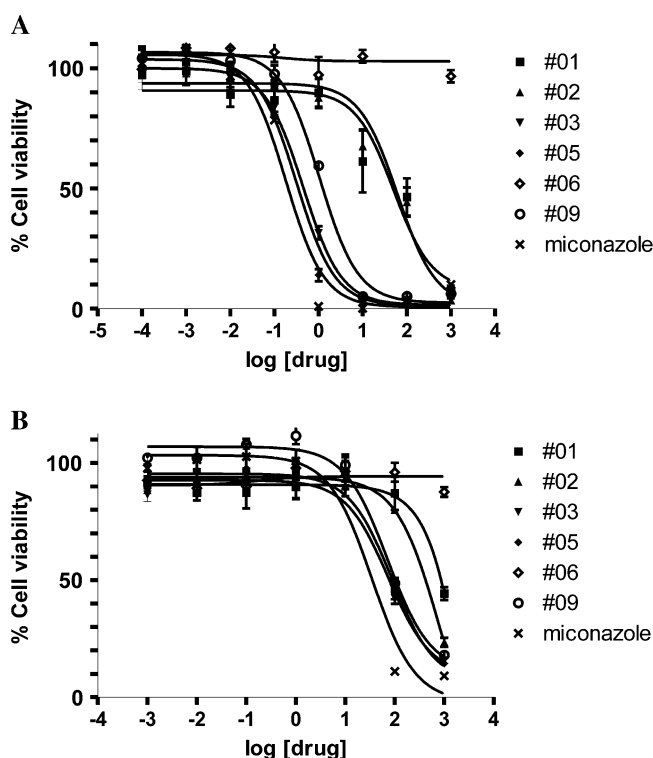


Fig. 5 Viability of LNCaP **(A)** and T47D **(B)** cells after treatment with the different synthesized compounds and control (XTT assay), log scale in μM.

Table II IC₅₀ Inhibition Values of Compounds Tested in LNCaP Prostate Cancer Cells and T47D Breast Cancer Cells

Compound	LNCaP IC ₅₀ (μM)	T47D IC ₅₀ (μM)
#01	50 ± 0.4	220.5 ± 0.9
#02	61 ± 0.2	198.2 ± 0.4
#03	0.44 ± 0.07	38.8 ± 0.2
#05	0.31 ± 0.05	44.5 ± 0.5
#06	No effect	No effect
#09	1.0 ± 0.1	49.1 ± 0.2
Miconazole	0.18 ± 0.02	40.9 ± 0.4

identified. Our experiments using the *Xenopus* oocyte expression system indicated that TH-1177 inhibited ⁴⁵Ca²⁺ influx derived from TRPV6 channel activity (Fig. 3). Moreover, it also inhibited TRPV5, the closest homolog of TRPV6 (Fig. 3 and Table I).

To develop more potent and selective inhibitors for TRPV6, we synthesized several compounds based on different chemical modifications of TH-1177 (Fig. 2) and checked their biological activities. Compound #03 was found to be the most potent and selective for TRPV6 (Fig. 3 and Table I), significantly inhibiting prostate and breast cancer cell proliferation (Fig. 5). Likewise, TRPV6-specific siRNAs were also able to reduce proliferation (Fig. 4), in agreement with previously published studies (34). In LNCaP cells, TRPV6 is the most abundant calcium entry channel and was expressed at levels around 45-fold higher than TRPV5 (37). The calcium channel expression data for T47D cells shows that TRPV6 is among the highest channels expressed (31). The expression of TRPV5 relative to TRPV6 is also dramatically lower. Therefore, we do not expect that TRPV5 plays a major role in calcium uptake in either LNCaP or T47D cells. It has been clearly demonstrated that TRPV6 controls calcium entry and proliferation in LNCaP cells (34). Similarly, in T47D cells, TRPV6 has been shown to play a major role in these processes (31).

The compounds that best inhibited TRPV6-mediated calcium uptake in oocytes were also the best inhibitors of cancer cell growth. A clear correlation was seen between the uptake inhibition and cell viability data. TH-1177 was previously shown to inhibit calcium entry into LNCaP cells, leading to reduced cell proliferation (35). These results strongly suggest that our compounds, especially #03, inhibit the calcium transport activity of endogenously expressed TRPV6 in prostate and breast cancer cells, and thus lead to lower proliferative ability. The benzhydryl-containing compounds #05 and #09 were also effective in reducing cancer cell proliferation. While the TRPV6 affinity of these compounds is lower than #03, it is high enough to produce a significant effect in the proliferation

assay. Our compounds might also be inhibiting other calcium entry channels present in the cancer cells. However, we believe the major effect is through TRPV6 inhibition.

Our experiments showed that the presence of an ester-bonded phenolic group in the synthesized compounds was critical for the inhibition against TRPV5 or TRPV6, since compound #06 was unable to inhibit either channel. The control compounds, econazole and miconazole, two other known inhibitors of TRPV5 and TRPV6, also contain this ester-bonded phenolic group at the center of the molecule (Fig. 2). This suggests that the benzhydryl or phenolic group is structurally important, probably reflecting the structure of binding pockets in TRPV5 and TRPV6 channels. Additionally, all the compounds with benzhydryl side groups were more TRPV5-selective, with compound #09 being the best inhibitor of TRPV5 (Table I). Interestingly, the most TRPV6-selective compound, #03, contained only an ester-linked chlorinated phenolic side group. Therefore, the nature of the ester-bonded side group seems to play a role in determining the channel selectivity of the compounds.

One can speculate that changing the π density of the aromatic groups by introducing different electron releasing/withdrawing groups could be a good strategy for further TRPV5-or TRPV6-targeted drug design, as described for a potassium channel (38). Although we investigated this in a limited way here, we can make a few noteworthy observations. Comparing compounds #01 and #02, it is evident that chlorinating the benzhydryl group produced a stronger TRPV-channel inhibitor, since both TRPV5 and TRPV6 were inhibited better by compound #01. Replacing the methoxyphenolic group (compound #01) with a chlorinated phenol (compound #09) further increased the effectiveness of the compound in inhibiting both channels. Compound #09, containing two chlorine-substituted phenolic groups, was the best inhibitor of TRPV5. When the chlorinated phenol was replaced with a dimethylaniline, the resulting compound (#05) became a slightly weaker TRPV5 inhibitor, but was the second best TRPV6 inhibitor. Compound #05 was the best benzhydryl-containing TRPV6 inhibitor tested. These compounds point the way to developing a larger group of inhibitors to probe deeper into the structure-activity relationships.

The selectivity and potency of compound #03 suggest that it could be a better compound for *in vivo* studies than TH-1177. TH-1177 treatment was shown to extend the average life span of mice carrying prostate tumors by up to 38% without toxicity (34). Therefore, compound #03 could be expected to perform as well or better. The systemic toxicity of compound #03 would be expected to be low because of its structural similarity to TH-1177. Studies of the systemic side effects of our compounds, as well as their effectiveness *in vivo*, are the next step.

CONCLUSION

We synthesized TRPV5 and TRPV6 inhibitors and identified one selective inhibitor for TRPV6. Several of our compounds were effective in inhibiting prostate and breast cancer cell proliferation. We propose that this growth inhibition may be caused by a reduction in TRPV6-mediated calcium uptake. Although we did not synthesize sufficient compounds to fully investigate structure-activity relationships, our study provides a starting point for the synthesis of further inhibitory compounds. Future studies will examine the effectiveness of our compounds in *in vivo* cancer models.

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